

Molecular MRI of apoptosis in atherosclerotic plaque by using a peptide-vectorized paramagnetic imaging probe

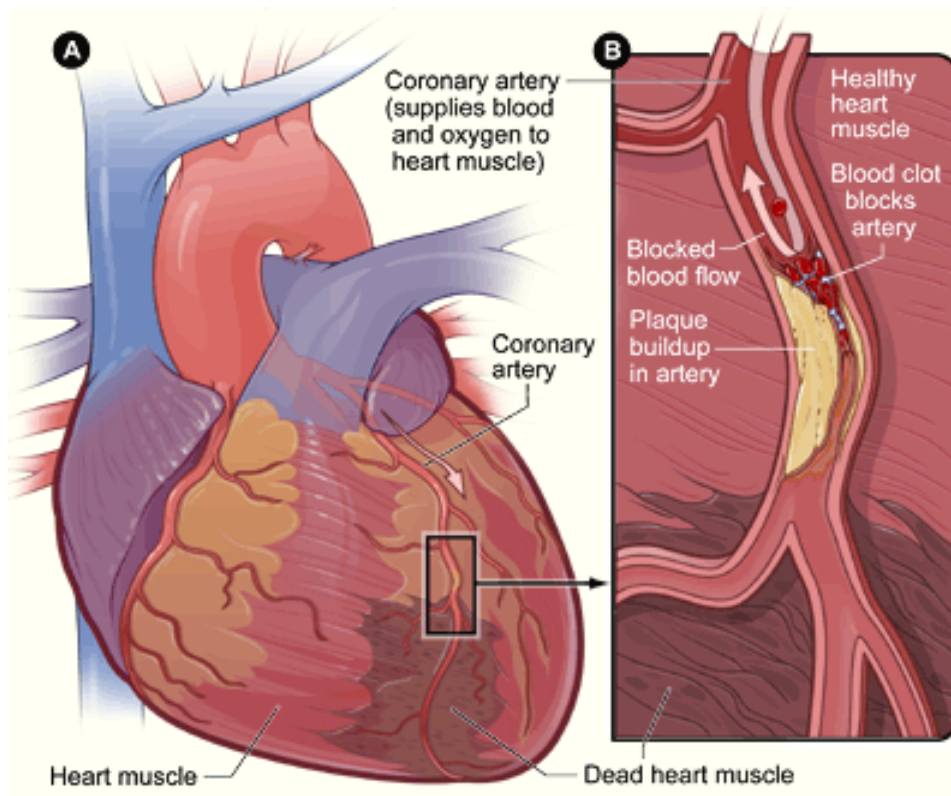


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<http://www.umh.ac.be/~nmrlab/>

Cardiovascular diseases: complications of atherosclerosis



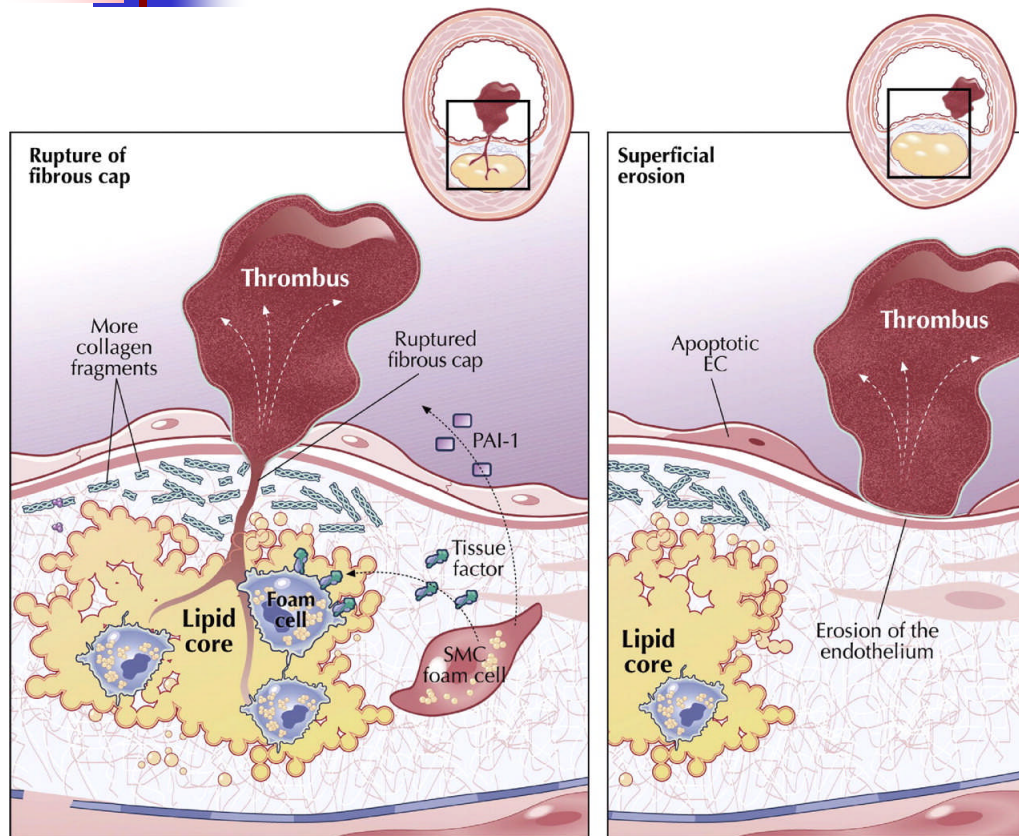
http://www.nhlbi.nih.gov/health/dci/Diseases/Cad/CAD_All.html

- Important progress in the therapy and prevention of cardiovascular diseases

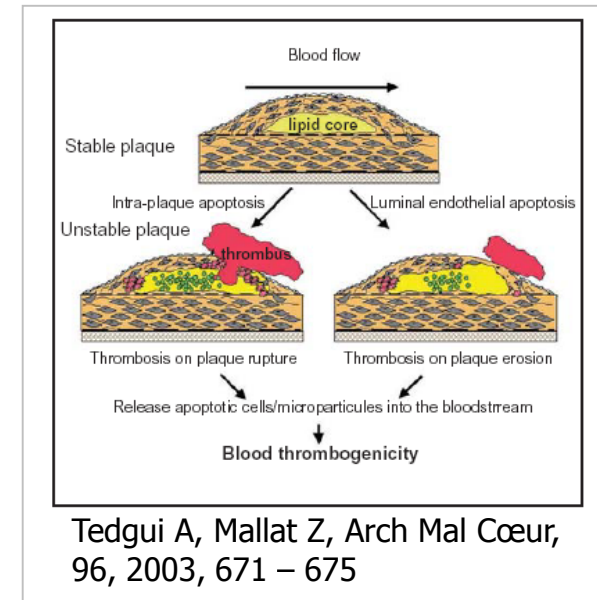


- Still, myocardial infarction and brain stroke → the main causes of death in Occidental countries

Vulnerable plaques, thrombosis and apoptosis

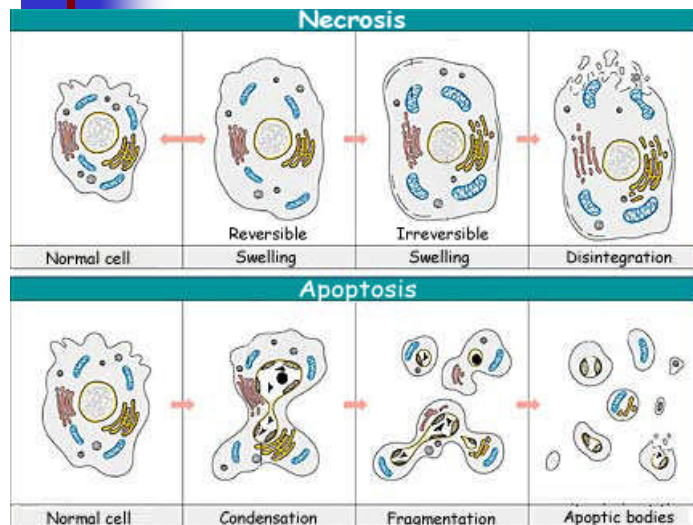


Libby, P. et al. J Am Coll Cardiol, 2006, 48, A33-A46



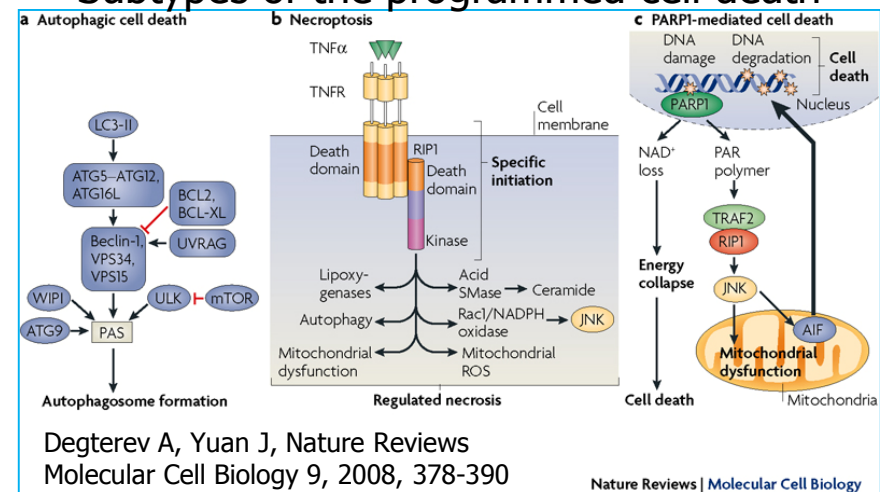
- Macrophages → MMPs → collagen degradation → weakening of the fibrous plaque → erosion and rupture of the fibrous plaque → thrombus
- Apoptosis of SMCs, macrophages, lymphocytes T → positive and negative effects

The mechanisms of cell death



http://thebrain.mcgill.ca/flash/capsules/outil_bleu17.html

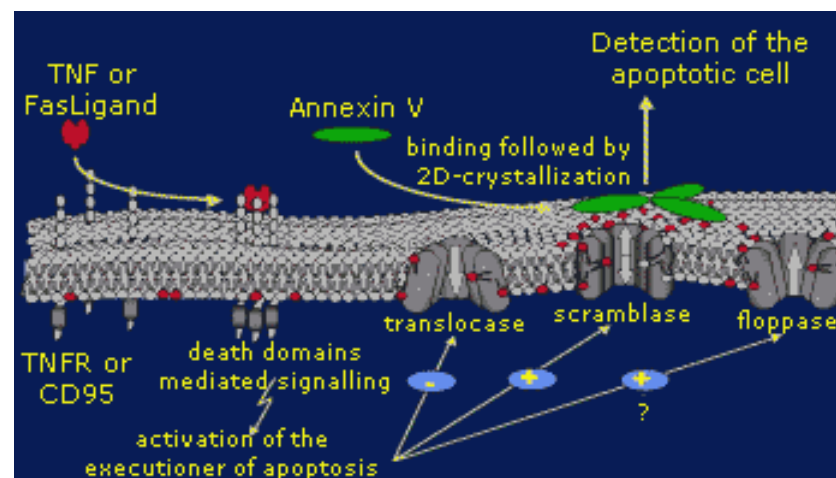
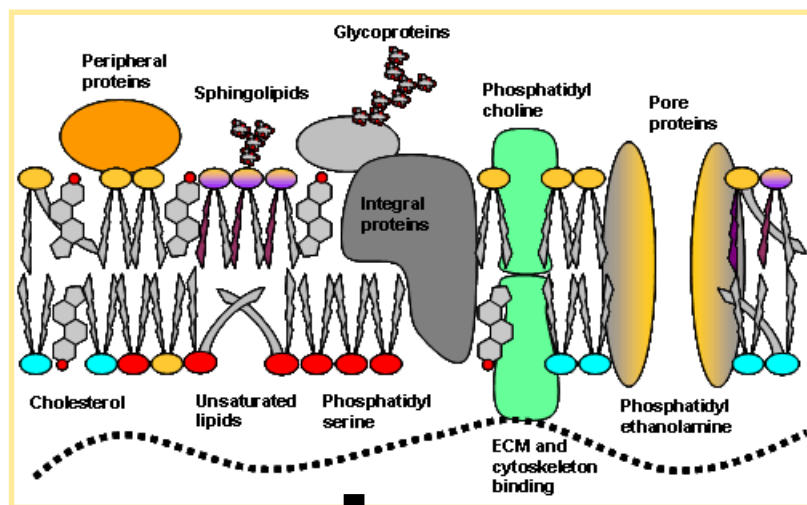
Subtypes of the programmed cell death



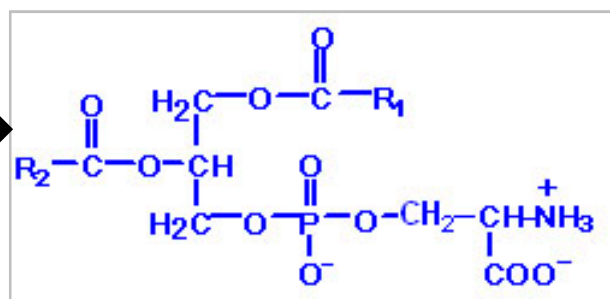
Apoptosis	Autophagy	Necroptosis	PARPtosis
<ul style="list-style-type: none"> - Chromatin fragmentation - Membrane blebbing - Apoptotic bodies - Caspase dependent - Early PS exposure 	<ul style="list-style-type: none"> - Expression of autophagy-related genes - Degradation of cell organelles - Accumulation of membrane-closed vesicles 	<ul style="list-style-type: none"> - Cell disintegration - Independent of caspases - Late PS exposure 	<ul style="list-style-type: none"> - Chromatin fragmentation independent of caspases - Energy collapse - Activation of PARP-1

AIM:

Search for peptide ligands that target apoptotic cells by a specific interaction with phosphatidylserine (PS)



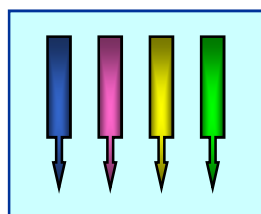
van Engeland M et al. Cytometry, 31, 1998, 1– 9



Phosphatidylserine

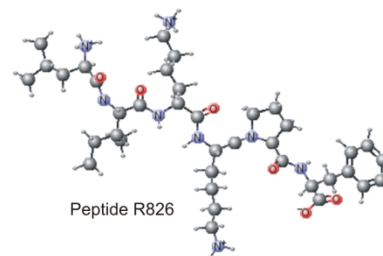
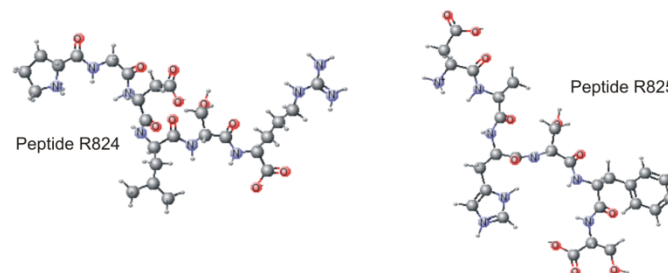
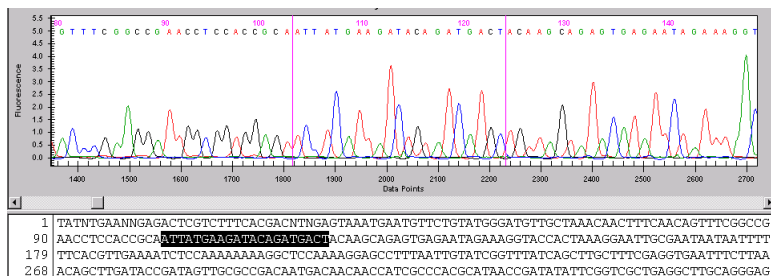
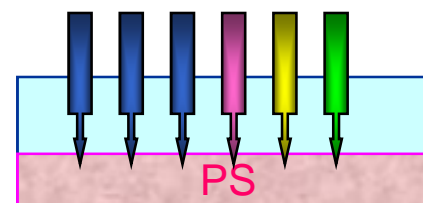
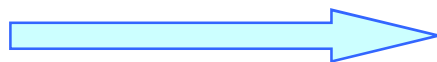
METHOD:

Phage display screening



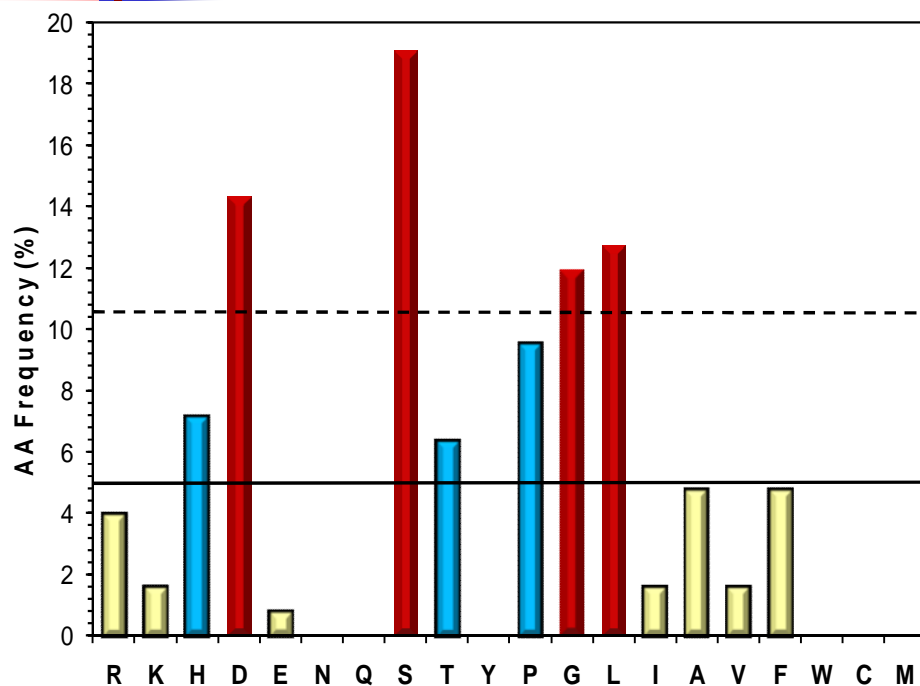
Library of linear
hexapeptides

3 rounds of panning



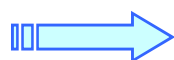
RESULTS:

Peptide sequence



3 first positions (N-terminal): Asp, His

3 last positions (C-terminal): Ser, Thr



Ionic or hydrogen binding with the polar head of PS

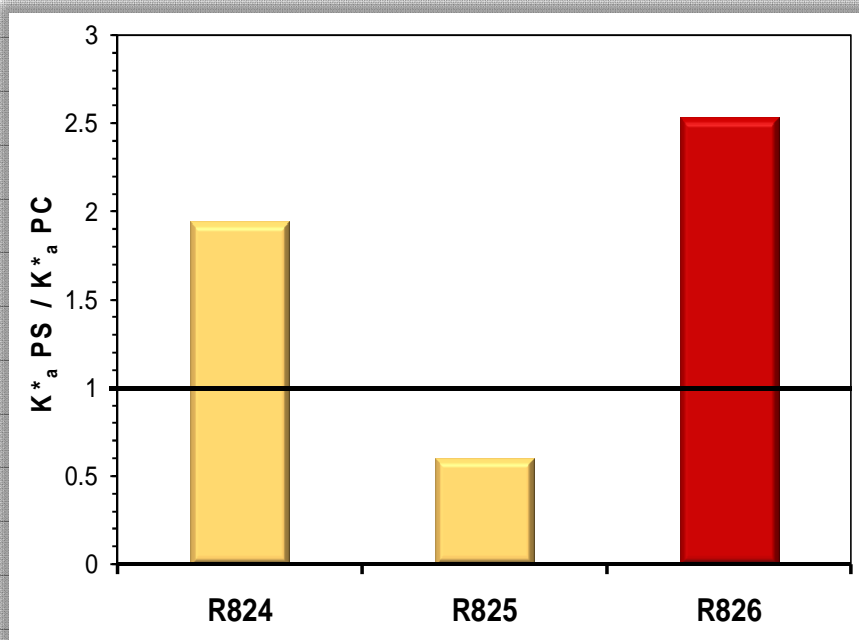
No clones	Homology
5	Low voltage-activated T-type calcium channel α-1 subunit
1	Tyrosine protein kinase pp60-c-src Neuronal pp60c-src
7	Matrix metalloproteinase 14 preprotein Matrix metalloproteinase 1, 9, 14
3	Transient rec. potential Ca^{2+} channel 6C Acyl-coenzyme A oxydase 2
1	Transient rec. potential Ca^{2+} channel 6C Fas antigen ligand
1	K^+ inwardly-rectifying channel K^+ large conductance pH-sensit. channel
1	Protein Tyr phosphatase 2C Alanine:glyoxylate aminotransferase 2
1	Apoptosis associated Tyr-kinase Ca^{2+} channel β-subunit
1	Transient receptor potential calcium channel 5 (TRPC5) Capacitative calcium entry channel 2

Theoretical biochemical parameters of the three candidate peptides R824, R825 and R826, respectively, as estimated by using ExPASy Proteomics Server, Proteomics and sequence analysis tools; LogP was calculated by using the ACDLabs 12.0 software

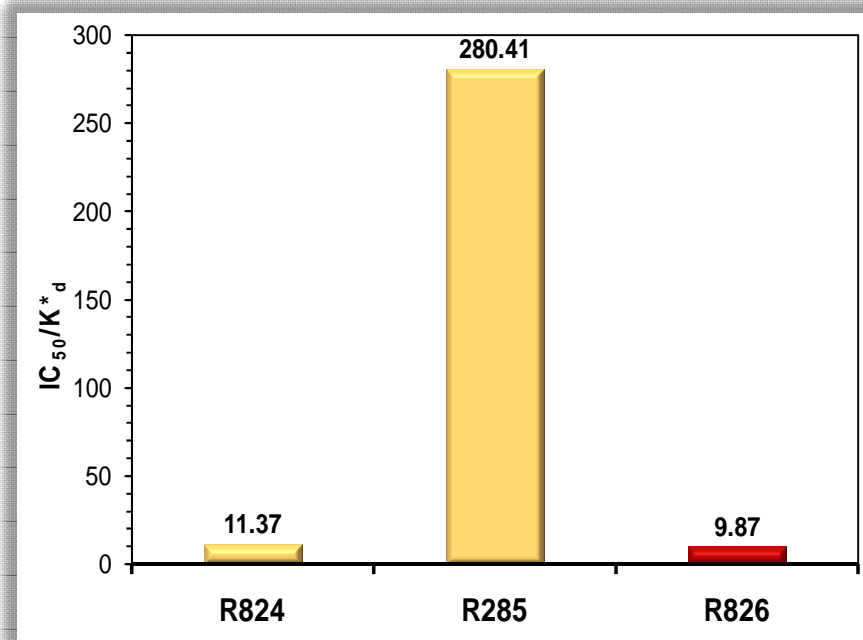
Parameter	R824	R825	R826
Half-life	>20 h	1.1 h	5.5 h
Instability index	40.43	-5.82	13.72
pI	6.27	5.08	10.00
LogP	-2.33 ± 0.86	-1.79 ± 0.88	2.51 ± 0.86
GRAVY	-1.167	-0.617	0.283
Aliphatic index	65	16.67	130

Half-life: theoretically estimated in mammalian reticulocytes *in vitro*; Instability index: when smaller than 40, the protein (or peptide) is predicted as stable; pI = Isoelectric point; LogP = Partition coefficient; GRAVY = Grand average of hydropathicity (predicts the hydrophobicity); Aliphatic index = the relative volume occupied by aliphatic side chains.

Specific affinity for PS of the selected peptides

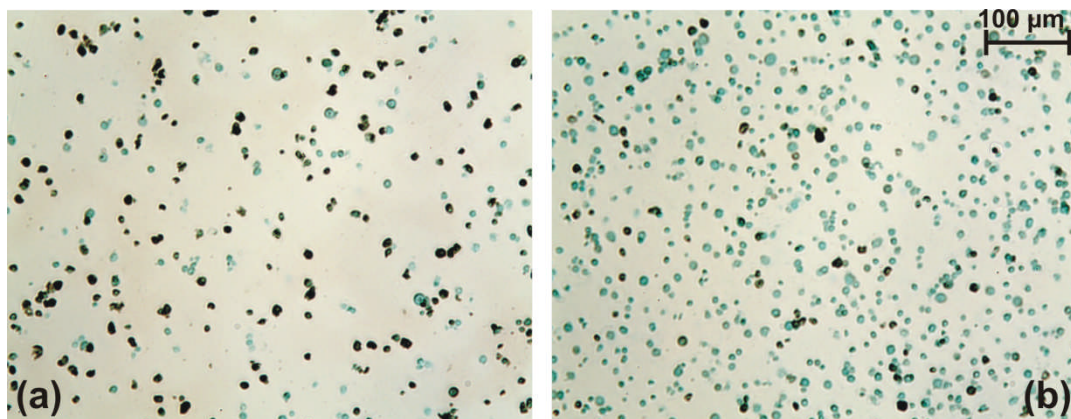


The ratio K_a for PS/ K_a for PC

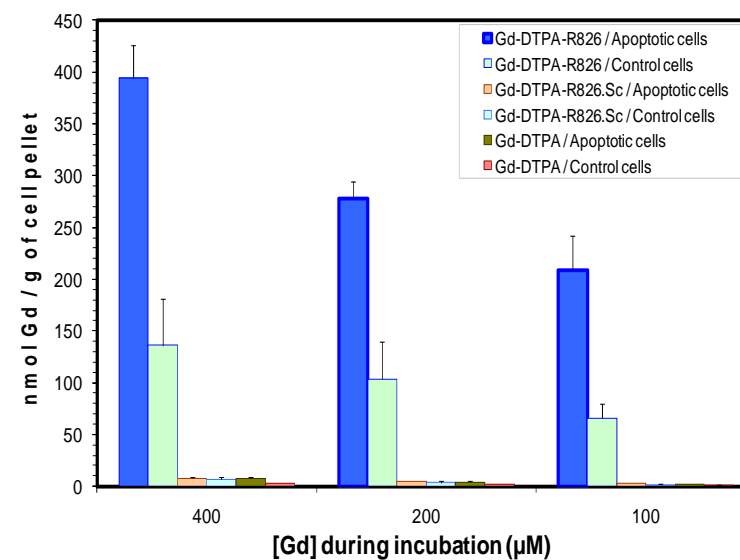
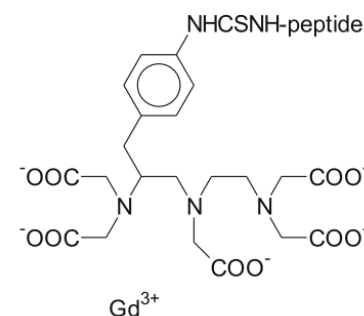


Ratio IC_{50}/K_d of PS-specific peptides.
The IC_{50} of R824, R825, and R826 was determined in competition with Annexin V

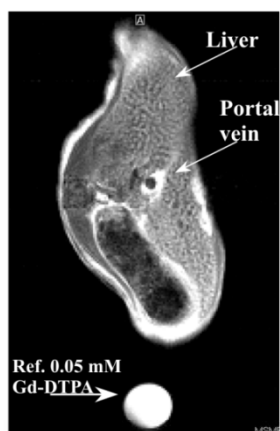
Binding of Gd-DTPA-g-R826 to apoptotic Jurkat cells as compared to various controls



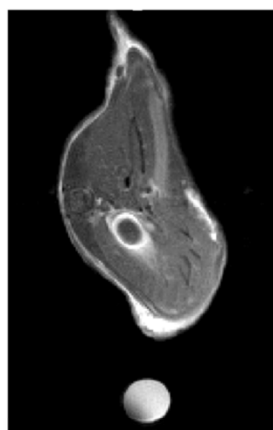
Apoptotic cells were stained (brown) in camptothecin treated **(a)** and control **(b)** samples with biotinylated Annexin V.



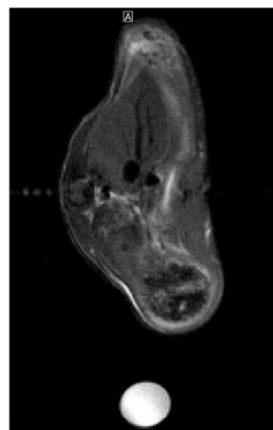
MRI (4.7T Bruker imaging system, T_1 -weighted MSME, TR/TE = 307.4/14.7 ms) of PS in mouse liver 30 min post contrast and immunohistochemistry of apoptotic cells



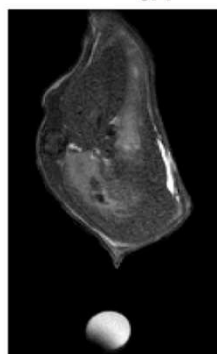
6A



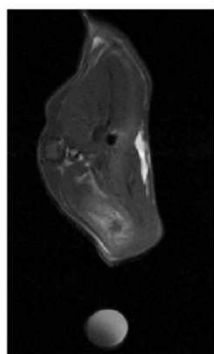
6B



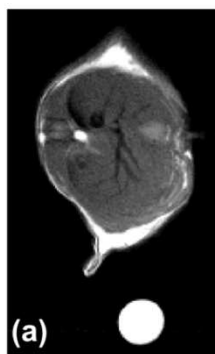
6C



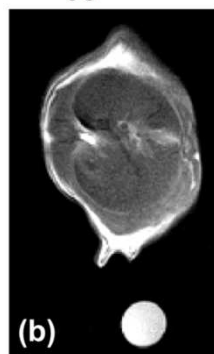
6D



6E

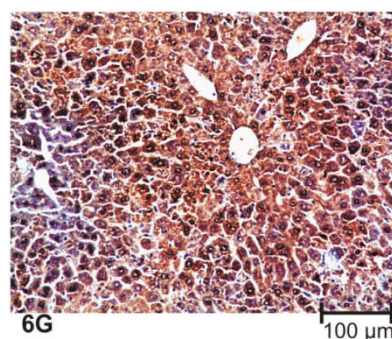


(a)

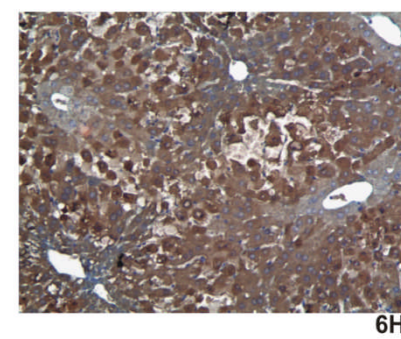


(b)

6F



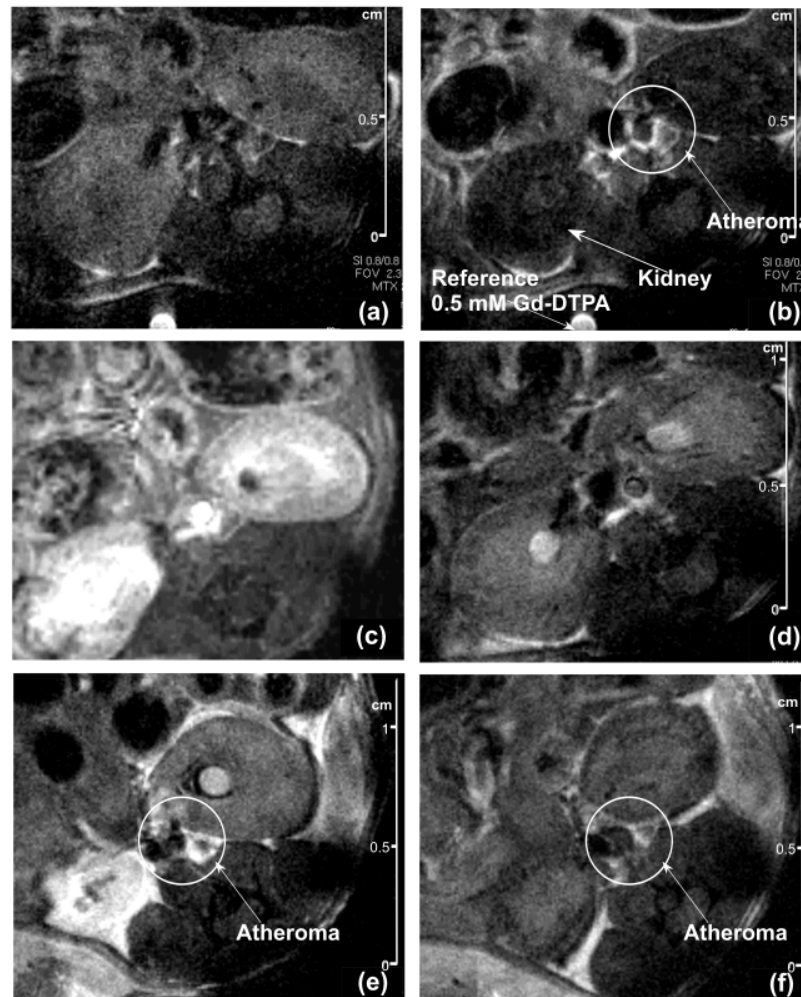
6G



6H

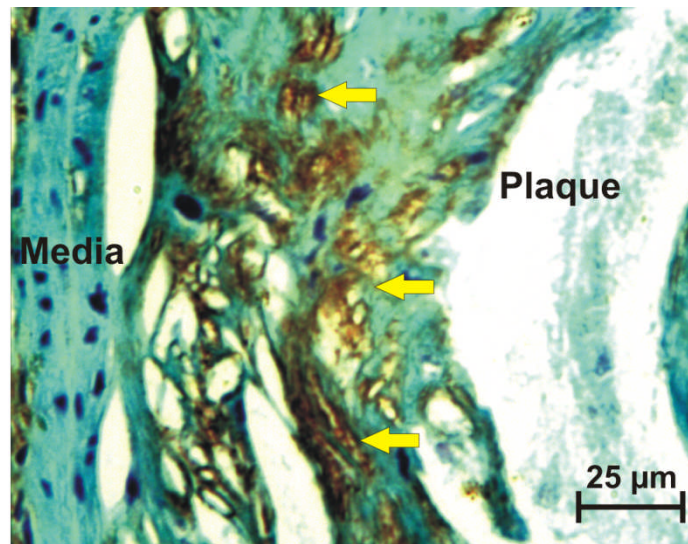
Apoptotic liver imaged with Gd-DTPA-g-R826 in the absence (**6A**) and in the presence of competitor R826 (**6B**) is compared to healthy liver (**6C**). Apoptotic (**6D**) and healthy (**6E**) liver imaged with Gd-DTPA. (**6F**) Apoptotic liver in pre-contrast (**a**) and post injection of Gd-DTPA-g-R826.Sc (**b**). Apoptotic cells immunostained (brown) with AnnV-Bt (**6G**) and anti-caspase-3 antibody (**6H**).

MR images (4.7T Bruker imaging system, RARE sequence, TR/TE = 1048.5/4 ms, spatial resolution = 90 μ m) of abdominal aorta in ApoE^{-/-} mice

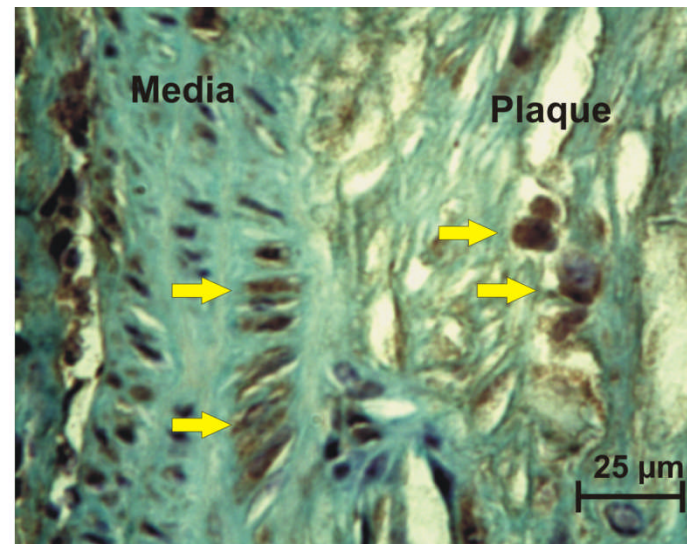


Axial slices of abdominal aorta are shown in pre-contrast **(a)** and ~30 min post Gd-DTPA-g-R826 **(b)**. They are compared to a TOF image **(c)** and to an image obtained post Gd-DTPA **(d)**. The comparison between Gd-DTPA-g-R826 **(e)** and Gd-DTPA-g-R826.Sc **(f)** is shown 60 min post-contrast. Images compared in **(a)–(d)** and those in **(e)–(f)** are located at the same level of abdominal aorta.

Immunostaining of apoptotic cells in atherosclerotic aorta of ApoE^{-/-} mice



(A)



(B)

Apoptotic cells were immunostained (brown) with biotinylated Annexin V **(A)** and with anti-caspase-3 antibody **(B)**.

Conclusions

- Peptide R826 → the most important PS-specific peptide
 - diagnosis of atherosclerotic disease and of other apoptosis-associated pathologies, such as cancer, ischemia, chronic inflammation, autoimmune disorders, transplant rejection, neurodegenerative disorders, and diabetes mellitus
 - cardiovascular diseases: apoptosis associated with loss of cardiomyocytes subsequent to myocardial infarction, atherosclerotic plaque instability, congestive heart failure and allograft rejection of the transplanted heart